



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 14 (2004) 5907-5911

Synthesis of analogs of (1,4)-3- and 5-imino oxazepane, thiazepane, and diazepane as inhibitors of nitric oxide synthases

K. Shankaran,^{a,*} Karla L. Donnelly,^a Shrenik K. Shah,^a Charles G. Caldwell,^a Ping Chen,^a William K. Hagmann,^a Malcolm MacCoss,^a John L. Humes,^b Stephen G. Pacholok,^b Theresa M. Kelly,^c Stephan K. Grant^c and Kenny K. Wong^c

^aDepartment of Medicinal Chemistry, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA
^bDepartment of Immunology and Inflammation Research, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA
^cDepartment of Enzymology, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA

Received 14 July 2004; revised 8 September 2004; accepted 8 September 2004 Available online 30 September 2004

Abstract—A series of 3- and 5-imino analogs from oxazepane, thiazepane, and diazepane was prepared and evaluated as inhibitors of human nitric oxide synthesis (NOS). The most potent iNOS inhibitor was the thiazepane analog **25** (IC₅₀ = $0.19 \,\mu\text{M}$). © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

The isozymes of nitric oxide synthase (NOS) produce nitric oxide (NO), a reactive inorganic radical gas, which plays central role in physiological functions in various mammalian tissues.^{2–5} All three NOS enzymes bring about a five electron oxidation of L-arginine to NO and L-citrulline, as shown in Figure 1. To date three isoforms of NOS enzymes have been identified. Of these, two isoforms are constitutive and are implicated in the cardiovascular and neuronal activities (eNOS and nNOS). The third isoform, which is linked with host

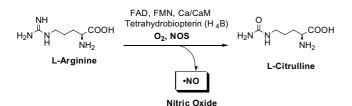


Figure 1. Nitric oxide biosynthesis pathway.

Keyword: NOS synthase.

defense, is induced by inflammatory cytokines and hence the name iNOS. Unwarranted production of nitric oxide by iNOS has been associated with tissue damage leading to inflammatory diseases such as arthritis and inflammatory bowl disease. Thus selective inhibition of iNOS may offer possible treatments for such diseases.

Efforts toward NOS inhibitors have led to amino acids as well as nonaminoacid based inhibitors, including aminoguanidines, isothioureas, amidines benzoxazolones, and 2-amino-pyridines, respectively. Previously, pyrrolidin-2-imines as inhibitors of NOS displaying selectivity to varying degrees has been reported (Fig. 2). In an obvious extension from the pyrrolidinyl and piperidinyl-2-imines, the analogous of 3-and 5-imino derivatives of oxazepane, thiazepane, and diazepane were prepared (Fig. 2). The synthesis and SAR of these classes of inhibitors are the subject of this report.

Figure 2.

[☆]See Ref. 1.

^{*}Corresponding author. Tel.: +1 732 594 3979; fax: +1 732 594 3007; e-mail: kothandaraman_shankaran@merck.com

2. Chemistry

The preparation of unsubstituted 5-imino analogs of oxazepane, thiazepane, and diazepane described herein are shown in Scheme 1. In an identical sequence of reactions both tetrahydropyran-4-one (X = O) and tetrahydrothiopyran-4-one (X = S) (1) were subjected to oximation followed by tosylation to give the desired tosylates (2). These tosylates (2) undergo smooth Beckmann rearrangement¹⁰ to afford the amides 3. The amides 3 were transformed to the thioethers 4 in two steps consisting of initial formation of the thioamide¹¹ from 3 that was followed by S-alkylation in the subsequent step. The thioethers 4 on reflux with ammonium chloride gave the amidines 5. The preparation of diazepane 8 took a different route starting from the known amide 6. Removal of the benzyl protecting group at nitrogen on 6 followed by reprotection gave 7. The transformation of 7 to the desired amidine 8 was accomplished in three steps that involve formation of iminoether, amidine, and then eventually N-Boc removal to afford 8 as shown below.

The synthesis of C-3 and C-6 substituted analogs of (1,4)-5-imino thiazepanes are shown in Scheme 2. The bis-allylester 9 was transformed to 11 in three steps

Scheme 1. Reagents and conditions: (a) NH₂OH, Py; (b) *n*-BuLi, TsCl, THF, -78 °C to rt; (c) dioxane, NEt₃, rt; (d) Lawesson's regent, Tol., 90 °C; (e) Me₃OBF₄, CH₂Cl₂; (f) NH₄Cl, EtOH, reflux; (g) Pd(OH)₂, H₂, EtOH, rt; (h) Boc₂O, CH₂Cl₂, rt; (i) HCl, EtOAc, rt.

Scheme 2. Reagents and conditions: (a) NaH, ether, allyl alcohol; (b) NaH, DMF, RI; (c) Pd(PPh₃)₄, THF, morpholine; (d) NH₂OH, Py; (e) *n*-BuLi, TsCl, THF, -78°C to rt; (f) dioxane, NEt₃, rt; (g) Lawesson's regent, Tol., 90°C; (h) Scheme 1 and steps e and f.

involving the Dieckmann cyclization ¹² of **9** to afford **10** that was followed by its alkylation and the eventual removal of allyloxycarbonyl moiety to furnish the tetrahydrothiopyran-4-one (**11**). Transformation of **11** to the oxime followed by the Beckmann rearrangement of the resultant oxime tosylate gave a mixture of two amides (**12**), that could not be separated. The mixture of **12** was converted to the thioamides **13** and **14**, at which point they could be separated by flash chromatography. The separated thioamides **13** and **14** were then transformed individually to their respective amidines **15** and **16** in a sequences of reaction similar to the conversion **3** to **5** described in Scheme 1.

The preparation of C-2 and C-7-methyl analogs of (1,4)-5-imino thiazepanes **19** and **20** were carried out as in Scheme 3. Oxidation of the β-ketoester **10** to the enone **17** was accomplished under active MnO₂ conditions as reported in the literature. The enone **17** underwent smooth Cu-catalyzed 1,4-addition of MeLi followed by Pd-mediated removal of the allyloxycarbonyl in subsequent step to give **18**. The conversion of **18** to the desired amidines **19** and **20** was analogous to Scheme 1.

The preparation of homochiral 3-(S)-propyl-(1,4)-5imino thiazepane **25** is shown in Scheme 4. The readily available N-Boc alcohol **21** undergoes facile aziridation¹⁴ under Mitsunobu protocol giving **22**. The nucleophilic ring opening of **22** by β -mercapto propionic acid gave the desired acid, that was subsequently deprotected

Scheme 3. Reagents and conditions: (a) active MnO₂, CHCl₃, rt; (b) MeLi, CuI–Me₂S, ether, -78°C to rt; (c) Pd(PPh₃)₄, THF, morpholine; (d) Scheme 1 and steps a–f.

Scheme 4. Reagents and conditions: (a) PPh $_3$, THF, DIAD, rt; (b) β -mercaptopropionic acid, Cs $_2$ CO $_3$, DMF, 60 °C; (c) EtOAc, HCl (g), rt; (d) EDC·HCl, DMF, NMM, rt; (e) Scheme 1 and steps d–f.

NC
$$\sim$$
 COOMe \longrightarrow a \longrightarrow C \longrightarrow NH \longrightarrow NH \longrightarrow 29 \longrightarrow S \longrightarrow C \longrightarrow NH \longrightarrow NH \longrightarrow 31 \longrightarrow 33 \longrightarrow 33

Scheme 5. Reagents and conditions: (a) RaNi, H₂, 1000 psi, 100 °C; (b) NaOEt, HSCH₂COOEt, EtOH; (c) Scheme 1.

to the amino acid 23.¹⁵ Transformation of 23 to the lactam 24 was accomplished using EDC, *N*-methylmorpholine in DMF. Conversion of 24 to the amidine 25 was analogous to Scheme 1. A similar sequence of reactions was in employment to transform 26 to the enantiomeric 3-(*R*)-propyl-(1,4)-5-imino thiazepane amidine 27.

The preparation of isomeric analogs of (1,4)-3-imino oxazepane (32) from 28 and the thiazepane (33) from 29 are shown in Scheme 5. This involved routine chemistry leading to the preparation of intermediate lactams 30 and 31 that was followed by steps given under Scheme 1 to give 32 and 33.

3. Results and discussion

The hydrochloride salts of synthesized (1,4)-3- and 5imino analogs were evaluated for NOS binding activity in an assay protocol that used recombinant versions of the human enzymes. ¹⁶ Results tabulated below describing the SAR for this class of inhibitors were compared with *N*-iminoethyl lysine (L-NIL) and primarily centered on the placement of substituents at all available position of the heterocycles. The IC₅₀ from the parent heterocycles are shown in Table 1. In the (1,4)-5-imine class, clearly the thiazepane analog 35 was relatively more potent and also had a better selectivity than the oxazepane analog 34, while the diazepane analog 8 was inactive $(IC_{50} > 50 \,\mu\text{M})$. The binding results for the (1,4)-3-imine analogs 32 and 33 are also displayed in Table 1. The relative comparison of the oxazepane analogs 34 and 32 suggested that the later was significantly more potent for the all three isoforms but neither displayed selectivity. Interestingly, the thiazepane analogs 35 and 33 were both equipotent and, unlike their oxazepane counterparts displayed slightly better selectivity. Since the selectivity data for both thiazepanes (33 and 35) appeared better, the effects of substituents on (1,4)-5-imino thiazepane (35) in order to enhance its potency and also to increase the selectivity for the iNOS over the other isoforms was explored. The assay results of these studies were compared with 35 and are shown in Table 2.

The thiazepane analog 36 with methyl at C-2 resulted in 2-folds loss of potency for iNOS compared to 35 and gain in the potency for other isoforms. The placement of methyl at C-3 (37) led to a gain in some potency for the all three isoforms. Analog 38 with a methyl at C-6 resulted in a substantial loss versus all isoforms, especially for the iNOS and eNOS, respectively. The presence of a methyl substituent at C-7 (39) gave an analog that was as potent as 35 or 36. Since the presence of methyl at C-3 generated more potent analog (37), further alterations were carried out at this position that led to the preparation of additional analogs bearing linear and branched alkyls, as in 40–45. The C-3 hydroxymethyl and bicyclic analogs 40 and 41 were unimpressive in that the former displayed no improvement compared to 37 while the later lost all activity. A 2–3-fold gain in the iNOS potency (compared to 37) was observed in the analogs 42–45. In general the gain in the selectivity over eNOS was substantial for these

Table 1. NOS inhibition by (1,4)-3- and 5-imino oxazepane, thiazepane, and diazepane^a

#	IC_{50} (μM)								
	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity eNOS/iNOS			
_	L-NIL	2.1	7.9	3.7	17.5	8.3			
8	H N N N NH	>50	>50	ND	>50	ND			
32	O N NH	0.16	0.45	2.8	0.16	1			
33	S N NH	1.2	10.5	8.8	2.8	2.3			
34	O N N NH	11.8	25	2.1	8.7	0.7			
35	S N NH	1.4	13.4	9.5	4.3	3			

^a IC₅₀s values are obtained from a 10 point titration using Sigma Plot, where each individual point is an average of duplicate determinations at that concentration.

^b The analogs in Table 1 were synthesized according to Schemes 1 and 5, ND = not determined, L-NIL = L-N-iminoethyl lysine.

Table 2. NOS inhibition by substituted (1,4)-5-imino thiazepanes^a

#	IC ₅₀ (μM)							
	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity eNOS/iNOS		
35	(S) N NH	1.4	13.4	9.5	4.3	3.0		
36	Me \ S \ N \ NH	3.5	4.1	1.1	0.83	0.24		
37	$Me \stackrel{S}{\underset{H}{\swarrow}}_{NH}$	0.81	5.0	6.1	1.8	2.2		
38	S — Me N NH	>50	>50	ND	>50	ND		
39	S Me NH NH	3.0	5.0	1.6	1	0.3		
40	HO SNH NH	1.1	6	5.3	2.7	2.4		
41	S Me	>50	>50	ND	>50	ND		
42	$Me \overset{S}{\underset{H}{\bigvee}}_{NH}^{S}$	0.31	1.1	3.7	0.59	2		
43	Me N N N N N N	0.29	50	172	2.8	10		
44	Me S N NH	0.32	60	186	1.9	6		
45	Ne NH NH	0.5	77	164	1.2	2.6		
25	$Me \xrightarrow{\prod_{\hat{H}}^{\hat{N}} N} NH$	0.19	1.0	5.2	0.44	2.3		
27	$Me \xrightarrow{H} \stackrel{S}{\underset{H}{\bigwedge}}_{NH}$	1.2	50	42	7.1	6		

^a IC₅₀s values are obtained from a 10 point titration using Sigma Plot, where each individual point is an average of duplicate determinations at that concentration

analogs while the selectivity over the nNOS was modest. As the *n*-propyl analog **43** was among the best, it was probed further to broaden the scope of these observations.

Since 43 was racemic, the chiral synthesis of enantiomers 25 and 27 were undertaken to see if an additional potency/selectivity are to be gained. Table 2 reveals that S-isomer (25) displayed 6-fold more potency for the iNOS compared to the (R)-isomer 27.

Paradoxically, the selectivity (over eNOS) data for the 25 was modest in comparison to 27. Interestingly, the

selectivity for neither compound matched that of the racemic analog 43 or other analogs displayed in Table 2.

In conclusion, limited SAR studies described herein established the (1,4)-5-imino thiazepanes as potent and selective iNOS inhibitors. We have also shown that iNOS potency can be improved by the placement of *n*-propyl substituent at C-3 of thiazepane and that the analog 25 was more potent and 27 was more selective. Our observations described herein is similar to an analogous studies conducted previously on these systems by Mooremann et al. The Nevertheless studies described herein when compared with the earlier observations suggest

^b In Table 2 the analogs were synthesized according to Schemes 2 and 3 (compounds **35–39**) and Scheme 4 (compounds **40–45** and including **25** and **27**), ND = not determined.

that the pyrrolidin-2-imine^{6h} or piperidin-2-imine^{6h,9} were significantly more potent and had a more balanced inhibitory profile than the analogs described herein.

References and notes

- The part work described herein was presented at the 212th ACS National Meeting, Washington, DC, 2000 (Abstract # 216).
- (a) Kerwin, J. F.; Lancaster, J. R.; Feldman, P. L. J. Med. Chem. 1995, 38, 4343; (b) Marletta, M. A. Cell 1994, 78, 927; (c) Feldman, P. L.; Griffith, O. W.; Stuehr, D. J. Chem. Eng. News 1993, 26; (d) Snyder, S. H.; Bredt, D. S. Sci. Am. 1992, 68.
- 3. Moncada, S.; Higgs, A. N. Engl. J. Med. 1993, 329, 2002.
- 4. White, K. A.; Marletta, M. A. Biochemistry 1992, 31, 6627.
- (a) Feldman, P. L.; Chi, S. Bioorg. Med. Chem. Lett. 1996,
 (b) Moore, W. M.; Webber, K. R.; Jerome, G. M.;
 Tjoeng, F. S.; Misko, T. P.; Currie, M. G. J. Med. Chem. 1994, 37, 3886; (c) Marletta, M. A. J. Med. Chem. 1994,
 37, 3886; (d) Narayanan, K.; Griffith, O. W. J. Med. Chem. 1994, 37, 885; (e) Olken, N. M.; Marletta, M. A. J. Med. Chem. 1992, 35, 3886.
- 6. (a) Cochran, F. R.; Selph, J.; Sherman, P. Med. Chem. Res. 1996, 16, 547; (b) Nussler, A. K.; Billiar, T. A. J. Leukocyte Biol. 1993, 54, 171; (c) Corbett, J. A.; Tilton, R. G.; Chang, K.; Hasan, K. S.; Ido, Y.; Wang, J. L.; Sweetland, M. A.; Lancaster, J. R.; Willamson, J. R.; McDaniel, M. L. Diabetes 1992, 41, 552; (d) Joly, G. A.; Ayres, M.; Chelly, F.; Kilbourn, R. G. Biochem. Biophys. Res. Commun. 1994, 199, 147; (e) Nakane, M.; Klinghofer, V.; Kuk, J. E.; Donnelly, J. L.; Budzik, G. P.; Pollock, J. S.; Basha, F.; Carter, G. E. Mol. Pharmacol. 1995, 47, 831; (f) Garvey, E. P.; Oplinger, J. A.; Tanoury, G. J.; Sherman, P. A.; Fowler, M.; Marshall, S.; Harmon, M. F.; Paith, J. E.; Furfine, E. S. J. Biol. Chem. 1994, 269, 26669; (g) Moore, W. M.; Webber, K. R.; Fok, K. F.; Jerome, G. M.; Connor, J. R.; Manning, P. T.; Wyatt, P. S.; Misko, T. P.; Tjoeng, F. S.; Currie, M. G. J. Med. Chem. 1996, 39, 669; (h) Shankaran, K.; Donnelly, K. L.; Shah, S. K.; Humes, J. L.; Pacholok, S.; Green, B. G.; Grant, S. K.; MacCoss, M. Bioorg. Med. Chem. Lett. 1997, 7, 2887; (i) Hagmann, W. K.; Caldwell, C. G.; Chen,

- P.; Durette, P.; Esser, C. K.; Lanza, T. J.; Kopka, I. E.; Guthikonda, R.; Shah, S. K.; Green, B. G.; Humes, J. L.; Kelly, T. M.; Luell, S.; Meurer, R.; Moore, V.; Pacholok, S. G.; Pavia, T.; Williams, H. R.; Wong, K. K. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 1975; (j) Ueda, S.; Terauchi, H.; Yano, A.; Ido, M.; Matsumoto, M.; Kawasaki, M. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 313.
- (a) Shankaran, K.; Donnelly, K. L.; Shah, S. K.; Guthikonda, R. N.; Humes, J. L.; Pacholok, S. G.; Grant, S. K.; MacCoss, M.; Wong, K. K. *Bioorg. Med. Chem. Lett.* 2004, 14, 4539; (b) Mooremann, A. E.; Metz, S.; Toth, M. V.; Moore, W. M.; Jerome, G.; Kornmeier, C.; Manning, P.; Hansen, D. W.; Ptzele, B. S.; Webber, R. K. *Bioorg. Med. Chem. Lett.* 2001, 11, 2651.
- 8. Hagen, T. J.; Bergmanis, A. A.; Kramer, S. W.; Fok, K. F.; Schmelzer, A. E.; Pitzele, B. S.; Swenton, L.; Jerome, G. M.; Kornmeier, C. M.; Moore, W. M.; Branson, L. F.; Connor, J. R.; Manning, P. T.; Currie, M. G.; Hallinan, E. A. J. Med. Chem. 1998, 41, 3675.
- 9. Guthikonda, R. N. National Medicinal Chemistry Symposium, 1996, Ann Arbor, MI, Abstract # 90.
- 10. Grob, C. A.; Ide, J. Helv. Chim. Acta 1974, 57, 2562.
- 11. Scheibye, S.; Pedersen, B. S.; Lawesson, S.-O. *Bull. Soc. Chim. Belg.* **1978**, *87*, 229.
- Casy, G.; Sutherland, A. G.; Taylor, R. J. K.; Urben, P. G. Synthesis 1989, 767.
- 13. Lane, S.; Quick, S. J.; Taylor, R. J. K. J. Chem. Soc., Perkin Trans. 1 1985, 893.
- (a) Pfister, J. R. Synthesis 1984, 969; (b) Mitsunobu, O. Synthesis 1981, 1; and also see Hughes, D. L. Org. React. 1992, 42, 335.
- 15. The crude acid 23 was taken as is to the next step to afford lactam 24 whose purification was facile.
- 16. The NOS inhibitory activity of the compound was determined by comparing the conversion of ³H-L-arginine to ³H-citrulline in the presence of inhibitor with control. The assay mixture containing 1μM of ³H-L-arginine, cofactors and the inhibitor or aqueous DMSO (control) was incubated for 30 min at room temperature. The reaction was quenched by adding a slurry of Dowex 50W-K8 resin to complex and remove the unreacted substrate. The concentration of the ³H-L-citrulline product in the supernatant fluid was determined on a scintillation counter. Under the assay conditions the production of L-citrulline was linear with time for the duration of the experiment.